



## Review

## Mpox infection in animals: A systematic review and meta-analysis



D. Katterine Bonilla-Aldana <sup>a,\*</sup>, Jorge Luis Bonilla-Aldana <sup>b</sup>, Juan R. Ulloque-Badaracco <sup>c</sup>, Ali Al-kassab-Córdova <sup>d</sup>, Enrique A. Hernandez-Bustamante <sup>e,f</sup>, Esteban A. Alarcon-Braga <sup>c</sup>, Vicente A. Benites-Zapata <sup>c</sup>, Cesar Copaja-Corzo <sup>g,k</sup>, Kenneth Silva-Cajaleon <sup>h</sup>, Alfonso J. Rodriguez-Morales <sup>h,i,j</sup>

<sup>a</sup> Research Unit, Universidad Continental, Huancayo, Peru

<sup>b</sup> Grupo de Investigación en Ciencias Animales Macagual, Universidad de La Amazonia, Florencia, Caquetá 180002, Colombia

<sup>c</sup> Escuela de Medicina, Universidad Peruana de Ciencias Aplicadas, Lima 15023, Peru

<sup>d</sup> Centro de Excelencia en Investigaciones Económicas y Sociales en Salud, Universidad San Ignacio de Loyola, Lima 15012, Peru

<sup>e</sup> Grupo Peruano de Investigación Epidemiológica, Unidad para la Generación y Síntesis de Evidencias en Salud, Universidad San Ignacio de Loyola, Lima 15012, Peru

<sup>f</sup> Sociedad Científica de Estudiantes de Medicina de la Universidad Nacional de Trujillo, Trujillo 13011, Peru

<sup>g</sup> Unidad de Investigación para la generación y síntesis de evidencias en salud, Universidad San Ignacio de Loyola, Lima 15012, Peru

<sup>h</sup> Faculties of Environmental and Health Sciences, Universidad Científica del Sur, Lima 4861, Peru

<sup>i</sup> Grupo de Investigación Biomedicina, Faculty of Medicine, Fundación Universitaria Autónoma de las Américas-Institución Universitaria Visión de las Américas, Pereira, 660003 Risaralda, Colombia

<sup>j</sup> Gilbert and Rose-Marie Chagoury School of Medicine, Lebanese American University, Beirut, Lebanon

<sup>k</sup> Servicio de infectología, Hospital Nacional Edgardo Rebagliati Martins, EsSalud, Lima 15072, Peru

## ARTICLE INFO

## Article history:

Received 5 February 2024

Received in revised form 26 March 2024

Accepted 17 April 2024

## Keywords:

Mpox

Poxviridae

Prevalence

Animals

Zoonotic

Transmission

## ABSTRACT

Mpox is a zoonotic disease that became epidemic in multiple countries in 2022. There is a lack of published systematic reviews on natural animal infection due to Mpox. We performed a systematic literature review with meta-analysis to assess animal Mpox prevalence. We performed a random-effects model meta-analysis to calculate the pooled prevalence and 95% confidence interval (95%CI) for prevalence studies. After the screening, 15 reports were selected for full-text assessment and included in qualitative and quantitative analyses. Ten reports assessed Mpox infection by molecular or serological tests ( $n = 2680$ ), yielding a pooled prevalence of 16.0% (95%CI: 3.0–29.0%) for non-human primates; 8.0% (95%CI: 4.0–12.0%) for rodents and 1.0% (95%CI: 0.0–3.0%) for shrews. Further studies in other animals are required to define the extent and importance of natural infection due to Mpox. These findings have implications for public human and animal health. OneHealth approach is critical for prevention and control.

© 2024 The Author(s). Published by Elsevier Ltd on behalf of King Saud Bin Abdulaziz University for Health Sciences. This is an open access article under the CC BY-NC-ND license (<http://creativecommons.org/licenses/by-nc-nd/4.0/>).

## Contents

Introduction	2
Methods	2
Information sources and search strategy	2
Eligibility criteria	2
Study selection and data extraction	3
Evaluation of studies quality and publication bias	3
Data synthesis and analysis	3
Results	4
Study selection and characteristics	4

\* Corresponding author.

E-mail address: [dbonilla@continental.edu.pe](mailto:dbonilla@continental.edu.pe) (D.K. Bonilla-Aldana).

Prevalence of Mpox in non-human primates . . . . .	4
Prevalence of Mpox in rodents . . . . .	5
Prevalence of Mpox in shrews . . . . .	5
Discussion . . . . .	5
Public health implications . . . . .	7
Limitations . . . . .	7
Ethical approval . . . . .	7
Funding source . . . . .	8
Declaration of Competing Interest . . . . .	8
Acknowledgements . . . . .	8
Declaration of competing interest . . . . .	8
Appendix A Supporting information . . . . .	8
References . . . . .	8

## Introduction

Mpox (MPX) is initially a zoonotic disease first described in 1958 [1] in non-human primates at the Stantens Serum Institut in Copenhagen, Denmark [2]. The MPX virus belongs to the Poxviridae family, the Chordopoxvirinae subfamily and the Orthopoxvirus genus. Genomically, they had two original lineages, the West African (WA) and the Congo Basin (Central Africa) clades, that differ genetically and in their virulence capacity, the latter being associated with more severe disease with higher mortality [3]. During the 2022 epidemics, evolutionary changes and a new proposed clade have been identified and reported [4,5].

In Europe, the first reported cases occurred in 2018 in Israel and the United Kingdom due to a traveller from Nigeria [6,7]. In May 2022, four positive cases of MPX were identified in the United Kingdom (UK), which had no link of origin or travel to endemic areas of the disease [8]. Other European countries reported positive cases in the coming months. Given the situation and possible additional transmission routes besides close contact, including sex, the World Health Organization declared mpox a public health emergency of international concern [9–12]. As of April 23, 2024, 95,340 cases were reported in 118 countries, with 184 confirmed deaths in 27 countries [13,14].

The transmission of MPX from infected animals to humans occurs mainly through direct contact with skin wounds, scabs, rashes, saliva, respiratory secretions, and body secretions through ingesting contaminated tissues and bites of infected animals [15]. In humans, transmission is associated with close contact with infected people with skin lesions or other areas, such as the mouth or genitals [16], with the virus entering through the injured skin, respiratory tract, and mucous membranes such as the oral, pharyngeal, ocular, genital, and anorectal [17]. The virus has been detected in sexual secretions, including semen [18,19]. In addition, the virus can be transmitted by fomites recently contaminated and vertically through the mother's placenta [13,20]. Then, Mpox has also been defined as a sexually transmitted infection (STI) and also associated with other STIs, including HIV, syphilis, and *Chlamydia* infection, among others [21,22]. Even in humans, a few cases of Mpox without detectable cutaneous/mucosal lesions have been described [23].

In humans, the disease has a prodromal period, which lasts from zero to five days with symptoms such as fever, headache, chills, myalgia, intense fatigue, and lymphadenopathy, mainly in the submandibular, postauricular, cervical, and inguinal areas. It should be noted that lymphadenopathy is a hallmark of MPX infection with other viruses such as measles, smallpox, and varicella [24,25]. Following fever, within one to three days, patients develop skin lesions frequently located on the mouth, face, palms of the hands and feet, genital area, conjunctiva, and perineal or perianal area [26,27]. Initially, rashes evolve from macules to papules, vesicles, pustules, and scabs that dry up and fall off, causing intense itching. Lesions in individuals vary from 10 to 150 and can last from 2 to 4 weeks until a

new layer of skin is generated. The sequelae of the disease are evidenced in hypopigmented or hyperpigmented varioliform scars [28].

Although the exact natural reservoir of the virus is not known, MPX has been isolated in multiple animal species so far, such as rodents (*Cricetomys* sp., *Jerbillo* sp. and *Chinchilla* sp.), non-human primates (*Cercocebus atys* and *Pan troglodytes verus*) and mammals (*Cynomys* sp.), among others [29–31]. In humans, the first case reported was in a 9-month-old boy in the Democratic Republic of the Congo in 1970, in which the family reported that they occasionally fed on monkeys [25]. Since then, cases have been confirmed and documented, especially in Africa (Cameroon, Ivory Coast, Liberia, Nigeria and Sierra Leone) in the 1970 s [32]. In 2003, a human outbreak occurred in the United States of America (USA) due to commercialising rodents imported from African countries that infected prairie dogs that were later commercialised [33,34].

Particularly with the 2022–2023 epidemics, multiple studies in humans and animals have been performed because there is concern about the role of animals as reservoirs, sources of infection, and receptors of infection from human sources, as confirmed initially in France in August 2022 [35,36].

According to the study, the observed range of the prevalence of Mpox infection in animals varies from 0% to 100%, according to the generated evidence. A systematic review with meta-analysis may help understand the risk and precisely know the global relative frequency of natural infection due to Mpox in animals. Unfortunately, to our knowledge, no other systemic reviews or meta-analyses have been published on this topic. The study's objective was to estimate the pooled prevalence of Mpox natural infection in animals based on available reports and observational studies.

## Methods

To achieve a thorough and accurate review, our work strictly followed the criteria outlined in the Preferred Reporting Items for Systematic Reviews and Meta-Analysis (PRISMA 2020) [37].

### Information sources and search strategy

On May 18, 2023, we conducted a literature search to find out the prevalence of animals diagnosed with Mpox. Using a controlled vocabulary thesaurus, the following databases were searched: PubMed, Scopus, Web of Science, and Ovid Medline. The search strategy was built using the Peer Review of Electronic Search Strategies (PRESS) Checklist [38]. No language nor geographic restrictions were applied. Please refer to [Supplementary Table S1](#) for the complete search strategy.

### Eligibility criteria

This review included cross-sectional/cohort studies that reported the prevalence of Mpox in either domestic or wild animals.

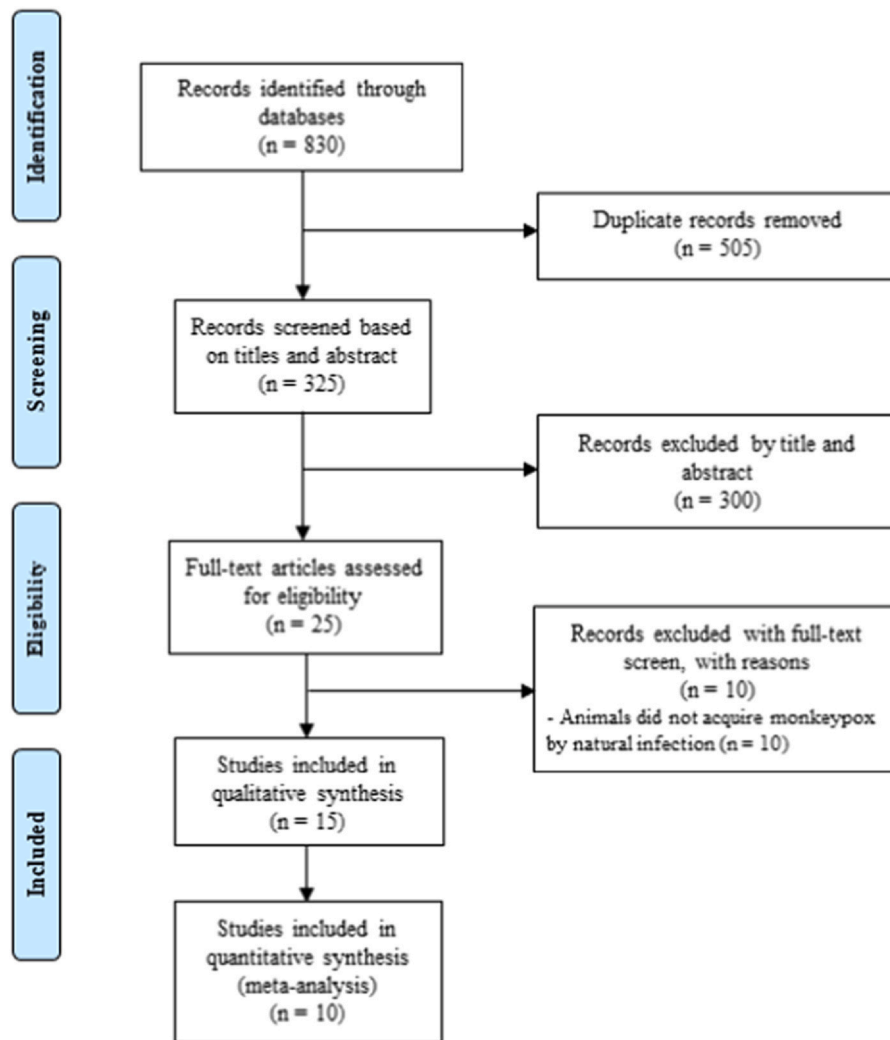


Fig. 1. Study selection and characteristics based on the PRISMA 2020 Standard for Systematic Reviews.

Additionally, we analyzed case reports and case series. All infections must have occurred naturally; owing to this, animals inoculated with the virus or that were part of experimental studies were not included. We also excluded articles that were 1) systematic reviews, 2) narrative reviews, 3) conference abstracts, 4) letters to the editor and 5) scoping reviews.

#### Study selection and data extraction

The findings from the literature search were transferred to the data management software 'Rayyan QCRI'. Four reviewers independently evaluated the titles and abstracts of the articles. Subsequently, each reviewer individually assessed the full text of the articles based on the predefined selection criteria. Any disagreement regarding the inclusion or exclusion of an article was resolved by reaching a consensus among the authors.

A data extraction sheet made in Google Sheets was used to collect the data from the selected articles. The following information was extracted: title, first author, study location, year, type of animal, and prevalence of Mpox infection.

#### Evaluation of studies quality and publication bias

For the risk of bias assessment, we used the Newcastle Ottawa scale adapted for Cross-sectional studies (NOS-CS), the Joanna Briggs

Institute Critical Appraisal Checklist for Case Reports and the Joanna Briggs Institute Critical Appraisal Checklist for Case series. In all three scales, a score of seven or more stars was considered as having a low risk of bias, whilst a score of fewer than seven stars indicated that the study being evaluated had a high risk of bias. The assessment was done independently by four reviewers. All disagreements were resolved by reaching a consensus among the authors.

The research team decided not to include the evaluation of publication bias. This decision was made in light of the shortcomings of Egger's tests and conventional funnel plots for assessing publication bias when used for proportional meta-analysis. As stated in previously published studies, this is based mainly on the following two factors. First, there needs to be more information to show that proportions fit these tests correctly. Second, it should be noted that the tests created to measure publication bias were initially based on the presumption that studies with favourable results were more likely to be published than those with unfavourable outcomes. There is no consensus on a favourable/unfavourable result in the case of proportions [39,40].

#### Data synthesis and analysis

The pooled prevalence rates with their corresponding 95% confidence intervals (95%CI) were calculated using STATA 16.0 with the package *metaprop*. For this quantitative analysis, we used a random

**Table 1**  
Characteristics of the included studies.

Author	Year	Country	Animals	Detection Method	Total Study	N	n (+)	%	
Tiee M et al.	2018	Central Africa	Rodent	PCR	1038	1038	93	9.0	
Orba Y et al.	2015	Zambia	Non-Human Primates	ELISA	978	188	4	2.1	
			Rodents	ELISA			259	38	14.7
			Shrews	ELISA			42	14	33.3
			Non-Human Primates	PCR			188	0	0.0
			Rodents	PCR			259	0	0.0
Doty J et al.	2017	Democratic Republic of the Congo	Shrews	PCR	346	42	0	0.0	
			Rodent	ELISA			262	6	2.3
			Shrews	ELISA			84	1	1.2
Hutson CL et al.	2007	USA	Rodent	PCR	254	254	33	13.0	
Breman JG et al.	1977	Ivory Coast and Mali	Non-Human Primates	PRNT	206	206	44	21.4	
Douglas KO et al.	2021	Barbados	Rodent	PCR	160	160	6	3.8	
Huntin Y et al.	2001	Democratic Republic of the Congo	Non-Human Primates	PRNT	54	6	0	0.0	
			Rodent	PRNT			44	13	29.5
			Pig	PRNT			4	1	25.0
Kulesh D et al.	2004	USA	Rodent	PCR	52	52	7	13.5	
Patrono L et al.	2020	Ivory Coast	Non-Human Primates	PCR	36	36	14	38.9	
Goldberg TL et al.	2008	Uganda	Non-Human Primates	ELISA	31	31	8	25.8	
Marennikova S et al.	1972	Democratic Republic of the Congo	Non-Human Primates	PRNT	9	9	2	22.2	
Langohr M et al.	2004	USA	Prairie Dog	PCR	1	1	1	100.0	
Guarner J et al.	2004	USA	Prairie Dog	PCR	2	2	2	100.0	
Radonić A et al.	2014	Ivory Coast	Non-Human Primates	PCR	1	1	1	100.0	
Seang S et al.	2022	USA	Dog	PCR	1	1	1	100.0	

effects model (Dersimonian and Laird method) and the 95%CI for each prevalence rate was calculated using the Clopper-Pearson method. The general meta-analysis was performed according to the kind of animal: non-human primates, shrews, and rodents. The Freeman-Tukey Double Arcsine Transformation was used as the variance stabiliser. Cochran's Q test and I<sup>2</sup> statistics were used to assess the heterogeneity between the studies. A p-value of less than 0.05 in Cochran's Q test revealed the presence of heterogeneity. For the I<sup>2</sup> statistic, values below 40% were deemed to represent mild heterogeneity, while values ranging from 41% to 60% indicated moderate heterogeneity and values exceeding 60% revealed substantial heterogeneity [39–41]. Subgroup analyses were conducted based on the detection method used (serological or molecular). For the sensitivity analysis, we excluded studies at high risk of bias. Case reports and case series were not included in meta-analyses.

## Results

### Study selection and characteristics

The search strategy returned 830 results. After eliminating duplicates, 325 articles were analysed, contrasting the selection criteria with the titles/abstracts. Then, 300 articles were evaluated in full text, and 15 studies were finally included in the review [29,34,35,42,43–52]. Fig. 1 illustrates the selection process with a PRISMA flowchart found in Fig. 1.

The characteristics of the included studies are summarised in Table 1. Of the 15 included studies, four were case reports, one a case series, and the remaining ten were cross-sectional studies; for a total of 16 datasets. The population of the included studies amounted to

2680 subjects, of which 477 were Non-Human Primates (NHP), 2069 were rodents, and 126 shrews, which were considered for meta-analysis. Additionally, four pigs, three prairie dogs and one domestic dog were analysed. The geographic location of the included studies was as follows: USA (5 studies), Democratic Republic of Congo (3 studies), Ivory Coast (3 studies but 1 of these studies was also conducted in Mali), Uganda (1 study), Zambia (1 study), Barbados (1 study), Central Africa (1 study). Regarding the detection method, six studies used a serological detection method (PRNT or ELISA) and ten studies used a molecular method (PCR).

As mentioned above, the risk of bias was assessed using three scales: the NOS-CS, the Joanna Briggs Institute Critical Appraisal Checklist for Case Reports, and the Joanna Briggs Institute Critical Appraisal Checklist for Case series. The evaluation identified that there were two studies at high risk of bias. The remaining studies were at low risk of bias. The table showing detailed information is found in Supplementary Table S2.

The quantitative synthesis of the prevalence of Mpox in dogs and pigs was not performed. In the case of pigs, the meta-analysis could not be performed because there was only 1 study assessing this population. In addition, quantitative synthesis was not conducted for dogs or prairie dogs in the case of reports due to the small sample size of only four animals, which could have introduced bias in the estimation.

The prevalence of Mpox varied according to the animals, and diagnostic tests were used across the different studies (Table 2).

### Prevalence of Mpox in non-human primates

The pooled prevalence of the studies evaluating the prevalence of Mpox in NHP was 16.0% (95%CI: 3.0–29.0%, five studies), showing

**Table 2**  
Summary of each meta-analysis results for the pool prevalence of Mpox among animals.

Animals and Diagnostic Methods <sup>a</sup>	Number of studies	N	Pool prevalence (%)	95%CI	I <sup>2</sup> ‡	p
Non-human primates (NHP)	5	467	<b>16.0</b>	3.0–29.0	93.30	< 0.01
NHP, only assessed by serological methods	4	431	10.0	0.0–29.0	93.78	< 0.01
Rodents	7	2069	<b>8.0</b>	4.0–12.0	90.66	< 0.01
Rodents, only assessed by serological methods	3	565	14.0	2.0–26.0	95.05	< 0.01
Rodents, only assessed by molecular methods	5	1763	7.0	3.0–12.0	87.66	< 0.01
Shrews	2	126	1.0	0.0–3.0	0.00	p > 0.05

95%CI = 95% confidence interval. ‡ I<sup>2</sup> index for the degree of heterogeneity.

<sup>a</sup> Some studies assessed simultaneous variables. Multiple studies assessed the prevalence by different methods.

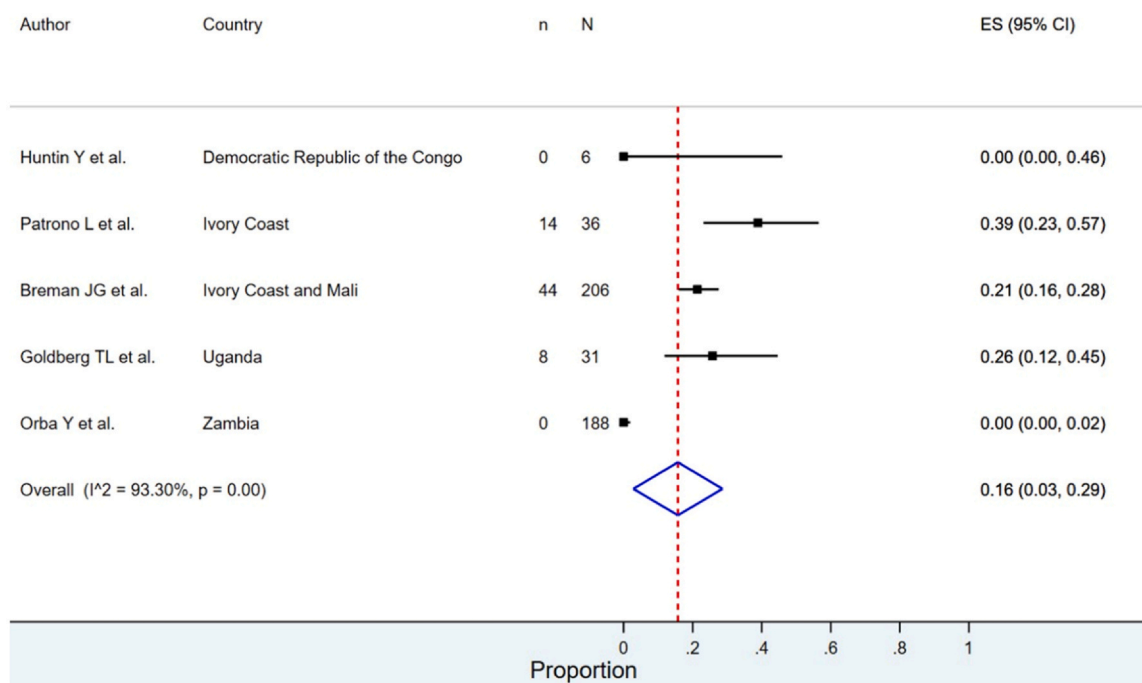


Fig. 2. Prevalence of Mpx in Non-Human Primates.

severe heterogeneity ( $I^2 = 93.30\%$ ) (Fig. 2). In the subgroup of studies that evaluated NHP with serological methods (Supplementary Fig. S1), the prevalence was 10.0% (95%CI: 0.0–29.0%). The sensitivity analysis (Supplementary Fig. S2) showed a prevalence of 27.0% (95%CI: 17.0–37.0%) with moderate heterogeneity ( $I^2 = 52.78\%$ ).

#### Prevalence of Mpx in rodents

The overall prevalence of Mpx in rodents was 8.0% (95%CI: 4.0–12.0%, seven studies) with severe heterogeneity ( $I^2 = 90.66\%$ ) (Fig. 3). In the subgroup of studies that evaluated rodents with serological methods (Supplementary Fig. S3), the prevalence was 14.0% (95%CI: 2.0–26.0%, three studies), and in the case of the studies that evaluated rodents with molecular methods (Supplementary Fig. S4), there was a prevalence of 7.0% (95%CI: 3.0–12.0%, five studies). The sensitivity analysis (Supplementary Fig. S5) showed a prevalence of 8.0% (95%CI: 3.0 - 12.0%) with severe heterogeneity ( $I^2 = 90.69\%$ ).

#### Prevalence of Mpx in shrews

The overall prevalence of Mpx in shrews was 1.0% (95%CI: 0.0 - 3.0%,  $I^2 = 0.0\%$ , two studies) (Fig. 4). No subgroup or sensitivity analysis was performed due to the few available studies.

## Discussion

The Mpx global outbreak has caused more than 95,000 cases in humans, with more than 180 associated deaths up to April 23, 2024, in 118 countries (111 that never reported mpox). Fortunately, the impact of this zoonotic disease has been entirely different compared to SARS-CoV-2/COVID-19 [53,54]. Mpx is not airborne nor transmitted by airdrops. The 2022–2023 outbreak has been associated with close contact, especially during human sexual activity [55,56], and the epidemic decreased after education and the use of vaccines against the disease [57,58].

Given the vast number of countries affected (> 110), in addition to the origins of the Mpx virus, it is critical, considering the extent of human-animal contact, to understand the potential risk derived

from the Mpx-infected humans to animals, especially outside Africa [3,35], but also there, where recently Mpx is also reemerging in some countries, as is the case the Democratic Republic of Congo (DRC) (almost 4,000 cases and more than 270 deaths in the last few months of 2023–2024) [59]. The main finding of the current meta-analysis indicated that around one in eight non-human primates (NHP) suspected and assessed by laboratory tests for Mpx infection was positive. That is a remarkable proportion of infection. Serological tests also found a high seroprevalence among NHP when the assessment was performed. Also, in this context, serological cross-reactions may occur, especially with other poxviridae that may infect animals. Some authors suggest that considering the Mpx evolution [4,5], the number of infected people and recent reports of environmental contamination [60–64], the possibility of Mpx transmission to animals can be expected more and more [65].

Indeed, in the current systematic review, the prevalence of Mpx was high in both molecular and serological analyses. However, more studies are needed, especially outside Africa, even after 2003 [52]. No studies about animals were published during 2022–2023, except a case report in France about the human-dog transmission of the virus [35]. Unfortunately, Mpx has been neglected for decades [66], and only recently, interest in the topic has increased with the 2022–2023 epidemics [67]. Even in Africa, where the disease has been endemic since the 1970s, there needs to be more knowledge regarding the disease burden in animals and humans [66]. It is of the utmost importance that more studies be performed on different groups of animals to approach the actual situation of natural infection in animals from this emerging family of poxviridae and understand its potential role as a reservoir and in transmission.

Since the 2003 epidemic in the USA, rodent animals have been on the radar of research-oriented efforts to describe the presence of Mpx infection, possible transmission and risk for humans [68,69]. However, early in the 2022–2023 epidemics, especially after the domestic dog case in France [35], the risk from humans to domestic animals and humans began to be assessed [70]. The increase in human Mpx cases raised concerns about the possibility of reverse zoonotic virus transmission from humans to animals. Consequently, in the UK, surveillance of pet animals living with individuals



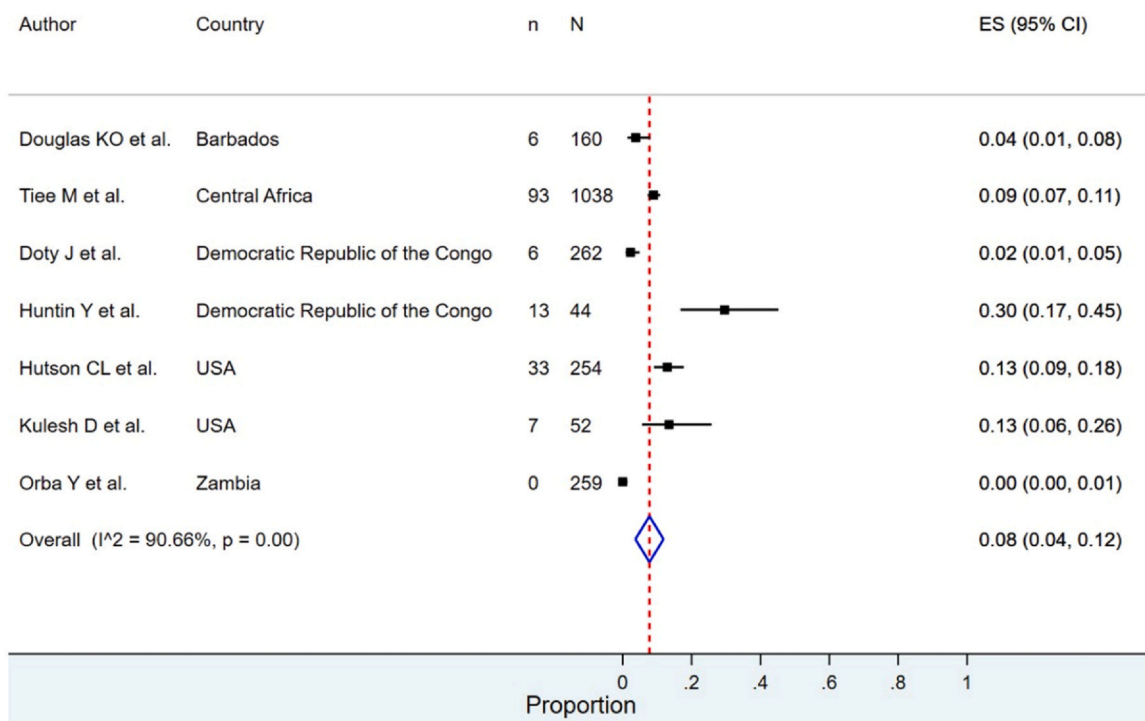


Fig. 3. Prevalence of Mpox in Rodents.

searching for the MPX virus started in May 2022 to assess the risk of this type of transmission. From June 1 to September 16, 2022, researchers collected data and observations during this time, showing that there have been no cases of animals with clinical signs suggestive of MPXV infection associated with confirmed human cases [70].

Understanding reverse zoonotic transmission of Mpox virus (MPXV) through pet animals is crucial for comprehending the potential risks of further transmission to humans and other animals. This information is valuable for informing the development and implementation of effective control measures to limit the spread of

the virus through this particular pathway. Surveillance of pets that cohabitate with individuals confirmed to have MPX is essential to this research. By monitoring and studying these pets, researchers can learn whether the virus can be transmitted from humans to animals and vice versa. By surveilling pets, scientists can assess the likelihood of reverse zoonotic transmission and quantify the risk it poses to humans and animals. This information is critical for understanding the potential for the virus to establish a reservoir in animal populations. If reverse zoonotic transmission is a significant risk, public health authorities can develop and refine measures to prevent such transmission. That might involve guidelines for

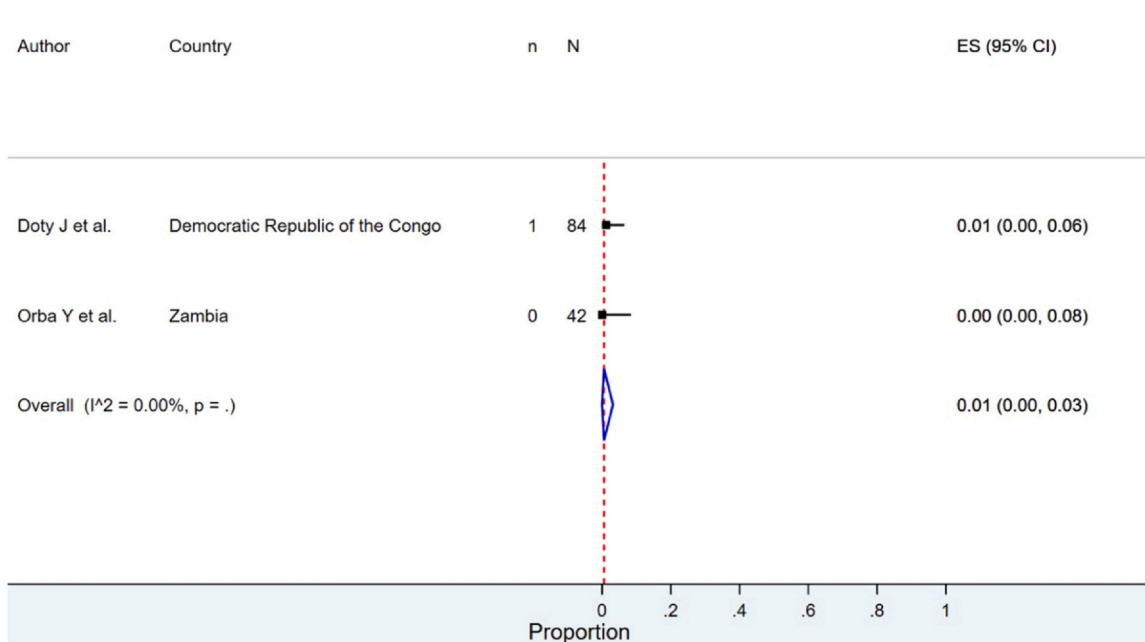


Fig. 4. Prevalence of Mpox in Shrews.

handling pets in households with confirmed cases of MPXV or recommendations for reducing close contact between infected individuals and their animals. Also, understanding how MPXV can potentially spread between humans and animals allows for proactive measures to prevent spillover events, where the virus jumps between species, is critical. That can help contain outbreaks and minimise the risk of broader transmission [71].

Addressing the potential for reverse zoonotic transmission requires a holistic approach to disease control. By considering the human-animal interface, control strategies can be more comprehensive and practical [55]. The results of animal surveillance can contribute to a better understanding of the dynamics of MPXV transmission, potential reservoir hosts, and the overall epidemiology of the disease. This knowledge is essential for formulating evidence-based strategies to manage and control human and animal cases, reducing the potential for further spread and cross-species transmission [70].

In the current meta-analysis, NHP, rodents and shrews showed a high prevalence of Mpox by different laboratory methods. However, most studies fail to indicate the clinical implications of MPXV infection in animals. Just a few of them described skin lesions in monkeys [1], as well as some in the Gambian pouched rats, showing lethargy, anorexia, weight loss, numerous skin lesions, vesicles on the tongue, and necrosis of the gingiva [72].

A study on prairie dogs also reported MPXV compromise in multiple organs and systems, including the liver, spleen, lungs, and heart, where the virus was detected [73]. Although more studies are needed on animals, those findings are consistent with those reported in humans during the extensive clinical studies performed during the 2022–2023 epidemics showing compromise of multiple organs and systems [74–77], especially in immunocompromised patients [78,79].

In the case of animals, although more animals would be involved, more studies need to address susceptibility to this virus. Domestic and wild animals must be investigated in different regions to understand better their potential role in disease and how many may act as animal reservoirs. So far, the number of animals assessed is globally limited, probably leading to overestimating the infected proportion in some animals but underestimating in others. Although individual case reports and small case series do not support prevalence figures, they may help as there is still a lack of studies on Mpox in animals.

In addition, many other animals may be included in similar systematic reviews in the future as soon as more studies are available, as is the case for non-human primates, especially rodents, which are susceptible to the infection caused by Mpox [3].

Considering the global epidemiology of Mpox, some countries with a high incidence of the disease have still not even published a single case of Mpox infection in these animals. That is the case of Brazil, Spain, Colombia, Mexico, and Peru, among others, which have reported more than 32% of all Mpox cases during the 2022–2023 epidemics. However, in the case of Brazil, there is an epidemiological report of a symptomatic 5-month-old dog in contact with a human confirmed case that also gets infected and confirmed [80]. Not much information regarding clinical evolution and outcome was available regarding the infected animals, but we could not find deaths associated with animals. These are uncommon in humans [14].

As observed, many of the most affected countries are in Latin America [81,82], a region severely affected by other sexually transmitted infections, such as HIV and syphilis [83–85], which also led to coinfections between Mpox virus and other sexually transmitted pathogens [86,87].

#### Public health implications

Many questions can be raised from the current cumulated evidence regarding the natural infection of Mpox in animals. However,

with the data available, there is an urgent need to consider its potential importance in transmission, interspecies, from human-to-animals, OneHealth perspectives that integrate human and animal health, when assessing cases occurring in domestic or wild environments, integrated surveillance and the need for increase regular testing among animals, beyond just research. There is a need to standardise molecular and serological tests for Mpox among animals [88], allowing these to be offered to the owners and increasing the diagnosis. At the same time, there is space for the discussion of more active surveillance instead of a passive report to OIE from the countries, promoting the search of animal cases among the cluster of human cases, as the UK has done [70]. Mpox deserves a comprehensive approach from the OneHealth approach [89,90]. More integration is still needed to increase our understanding of transmission, risks and consequences of this emerging poxviridae disease. Finally, now that vaccination is being used more widely than in the past in humans [57], there is also the question of its use in animals. So far, there has not been an approved orthopoxvirus vaccine for animals in the USA and other countries. However, developments may have occurred since then. It is now more apparent than ever that we need a better and more proactive approach. One possibility is to eliminate the threat of spillover before it occurs using vaccines capable of autonomously spreading through wild animal reservoirs. We are poised to develop self-disseminating vaccines targeting many human pathogens, including Mpox. However, essential decisions remain about how they can be most effectively designed and used to target pathogens with a high risk of spillover and emergence [91].

#### Limitations

In this study, we did not differentiate studies assessing the prevalence of screenings and the prevalence of outbreaks in closed groups of animals in detail. Due to an inherent limitation of cross-sectional/survey studies included in this review, the estimates in the primary studies could change from time to time. On the other hand, the merit of observational studies is that they can assess health problems in a natural setting, reflecting real-life situations. Some studies were performed in the context of epidemics but needed to be specified. Nevertheless, then, the data for it is still limited. Subsequently, additional analysis should be performed using more available and specific studies. In future assessments, it would be good to have a clear distinction between studies testing randomly in the wild population where animals are tested or when found dead and animals in commercial situations (tested when there appears to be an outbreak) or companion animals (tested often concerning positive PCR results of their owners) all of which has a high impact on the chances of finding a PCR-positive result. Doubtless, a more in-depth analysis of this would be interesting. Likewise, substantial heterogeneity may exist even with several stratifications of analysis of studies with more homogenous subgroups (e.g., g by species, diagnostic methods, or animal categories). True heterogeneity is expected in prevalence estimates due to variations in the time and place of included studies. Therefore, a high  $I^2$  does not necessarily mean inconsistent data (Barker et al., 2021). Hence, interpretation of the findings should be undertaken with caution.

Finally, a thorough review of Mpox natural infections in animal species is needed, emphasising how to interpret the findings of other authors in future systematic reviews and meta-analyses.

#### Ethical approval

Approval was not required.

## Funding source

The Universidad Continental, Peru, supported the publication fees of this article. This research was granted by the Universidad Peruana de Ciencias Aplicadas through the ExPost 2024 bonus.

## Declaration of Competing Interest

All authors report no potential conflicts.

## Acknowledgements

None.

## Declaration of competing interest

All authors report no potential conflicts.

## Appendix A. Supporting information

Supplementary data associated with this article can be found in the online version at doi:10.1016/j.jiph.2024.04.015.

## References

- Magnus Pv, Andersen EK, Petersen KB, Birch-Andersen A. A pox-like disease in cynomolgus monkeys. *Acta Pathol Microbiol Scand* 1959;46(2):156–76.
- Franco LAO, Moreno-Samper D, Chaparro-Mérida NA. Viruela del simio. *Rev Chil De Infectol* 2022;39:457–66.
- Bonilla-Aldana DK, Rodríguez-Morales AJ. Is monkeypox another reemerging viral zoonosis with many animal hosts yet to be defined? *Vet Q* 2022;42(1):148–50.
- Luna N, Ramirez AL, Munoz M, Ballesteros N, Patino LH, Castaneda SA, et al. Phylogenomic analysis of the monkeypox virus (mpxv) 2022 outbreak: emergence of a novel viral lineage? *Travel Med Infect Dis* 2022;49:102402.
- Luna N, Munoz M, Bonilla-Aldana DK, Patino LH, Kasminskaya Y, Paniz-Mondolfi A, et al. Monkeypox virus (mpxv) genomics: a mutational and phylogenomic analyses of b.1 lineages. *Travel Med Infect Dis* 2023;52:102551.
- Vaughan A, Aarons E, Astbury J, Balasegaram S, Beadsworth M, Beck CR, et al. Two cases of monkeypox imported to the united kingdom, september 2018. *Eur Surveill* 2018;23(38).
- Erez N, Achdout H, Milrot E, Schwartz Y, Wiener-Well Y, Paran N, et al. Diagnosis of imported monkeypox, israel, 2018. *Emerg Infect Dis* 2019;25(5):980–3.
- Vivancos R, Anderson C, Blomquist P, Balasegaram S, Bell A, Bishop L, et al. Community transmission of monkeypox in the united kingdom, april to may 2022. *Eur Surveill* 2022;27(22).
- Multi-country monkeypox outbreak: Situation update <https://www.who.int/emergencies/disease-outbreak-news/item/2022-don393>.
- Sah R, Reda A, Lashin BI, Mohanty A, Abdelaal A, Rodríguez-Morales AJ. Public health emergencies of international concern in the 21st century. *Ann Med Surg* (2012) 2022;81:104417.
- Leon-Figueroa DA, Bonilla-Aldana DK, Pachar M, Romani L, Saldana-Cumpa HM, Anchay-Zuloeta C, et al. The never-ending global emergence of viral zoonoses after covid-19? The rising concern of monkeypox in europe, north america and beyond. *Travel Med Infect Dis* 2022;49:102362.
- Sah R, Reda A, Abdelaal A, Mohanty A, Siddiq A, Alshahrani NZ, et al. A potential monkeypox pandemic: are we making the same mistakes as covid-19? *N Microbes N Infect* 2022;49:101030.
- Mpox (monkeypox) <https://www.who.int/news-room/fact-sheets/detail/monkeypox>.
- Sah R, Mohanty A, Abdelaal A, Reda A, Rodríguez-Morales AJ, Henao-Martinez AF. First monkeypox deaths outside africa: no room for complacency. *Ther Adv Infect Dis* 2022;9. 20499361221124027.
- Brown K, Leggat PA. Human monkeypox: current state of knowledge and implications for the future. *Trop Med Infect Dis* 2016;1(1).
- Rodríguez-Morales AJ, Lopardo G. Monkeypox: another sexually transmitted infection? *Pathog (Basel, Switz)* 2022;11(7).
- Ortiz-Martínez Y, Rodríguez-Morales AJ, Franco-Paredes C, Chastain DB, Gharamti AA, Vargas Barahona L, et al. Monkeypox - a description of the clinical progression of skin lesions: A case report from colorado, USA. *Ther Adv Infect Dis* 2022;9. 20499361221117726.
- Reda A, Abdelaal A, Brakat AM, Lashin BI, Abouelkheir M, Abdelazeem B, et al. Monkeypox viral detection in semen specimens of confirmed cases: a systematic review and meta-analysis. *J Med Virol* 2023;95(1):e28250.
- Barboza JJ, Leon-Figueroa DA, Saldana-Cumpa HM, Valladares-Garrido MJ, Moreno-Ramos E, Sah R, et al. Virus identification for monkeypox in human seminal fluid samples: A systematic review. *Trop Med Infect Dis* 2023;8(3).
- Dashraath P, Nielsen-Saines K, Mattar C, Musso D, Tambyah P, Baud D. Guidelines for pregnant individuals with monkeypox virus exposure. *Lancet* 2022;400(10345):21–2.
- Ciccarese G, Di Biagio A, Bruzzone B, Guadagno A, Taramasso L, Oddenino G, et al. Monkeypox outbreak in genoa, italy: Clinical, laboratory, histopathologic features, management, and outcome of the infected patients. *J Med Virol* 2023;95(2):e28560.
- Labate L, Brucci G, Ciccarese G, Bruzzone B, Ricucci V, Stefanelli F, et al. Nasal monkeypox virus infection successfully treated with cidofovir in a patient newly diagnosed with hiv. *Int J STD AIDS* 2023;34(3):208–10.
- Ciccarese G, Brucci G, Di Biagio A, Drago F, Ogliastrò M, Caccianotti B, et al. Two cases of monkeypox virus infection without detectable cutaneous/mucosal lesions. *Travel Med Infect Dis* 2023;54:102605.
- Johri N, Kumar D, Nagar P, Maurya A, Vengat M, Jain P. Clinical manifestations of human monkeypox infection and implications for outbreak strategy. *Health Sci Rev (Oxf)* 2022;5:100055.
- Singhal T, Kabra SK, Lodha R. Monkeypox: A review. *Indian J Pediatr* 2022;89(10):955–60.
- Petersen E, Kantele A, Koopmans M, Asogun D, Yinka-Ogunleye A, Ihekweazu C, et al. Human monkeypox: Epidemiologic and clinical characteristics, diagnosis, and prevention. *Infect Dis Clin North Am* 2019;33(4):1027–43.
- Thornhill JP, Barkati S, Walmsley S, Rockstroh J, Antinori A, Harrison LB, et al. Monkeypox virus infection in humans across 16 countries - april-june 2022. *N Engl J Med* 2022;387(8):679–91.
- Catala A, Riera J, Fuertes I. Mpox - formerly monkey pox - in dermatology: a review of epidemiologic features, clinical presentation, diagnosis, and treatment. *Actas Dermosifiliogr* 2023;114(4):318–26.
- Patrono LV, Pleh K, Samuni L, Ulrich M, Rothemeier C, Sachse A, et al. Monkeypox virus emergence in wild chimpanzees reveals distinct clinical outcomes and viral diversity. *Nat Microbiol* 2020;5(7):955–65.
- Kulesh DA, Loveless BM, Norwood D, Garrison J, Whitehouse CA, Hartmann C, et al. Monkeypox virus detection in rodents using real-time 3'-minor groove binder tagman assays on the roche lightcycler. *Lab Invest* 2004;84(9):1200–8.
- Doshi RH, Guagliardo SAJ, Doty JB, Babeaux AD, Matheny A, Burgado J, et al. Epidemiologic and ecologic investigations of monkeypox, likouala department, republic of the congo, 2017. *Emerg Infect Dis* 2019;25(2):281–9.
- Harapan H, Ophinni Y, Megawati D, Frediansyah A, Mamada SS, Salampe M, et al. Monkeypox: a comprehensive review. *Viruses* 2022;14(10).
- Guarner J, Johnson BJ, Paddock CD, Shieh WJ, Goldsmith CS, Reynolds MG, et al. Monkeypox transmission and pathogenesis in prairie dogs. *Emerg Infect Dis* 2004;10(3):426–31.
- Langohr IM, Stevenson GW, Thacker HL, Regnery RL. Extensive lesions of monkeypox in a prairie dog (cynomys sp). *Vet Pathol* 2004;41(6):702–7.
- Seang S, Burrell S, Todesco E, Leducq V, Monsel G, Le Pluart D, et al. Evidence of human-to-dog transmission of monkeypox virus. *Lancet* 2022;400(10353):658–9.
- Sykes JE. A call for more evidence documenting human-to-dog transmission of monkeypox virus. *Lancet* 2022;400(10357):993.
- Page MJ, McKenzie JE, Bossuyt PM, Boutron I, Hoffmann TC, Mulrow CD, et al. The prisma 2020 statement: an updated guideline for reporting systematic reviews. *BMJ (Clin Res Ed)* 2021;372:n71.
- McGowan J, Sampson M, Salzwedel DM, Cogo E, Foerster V, Lefebvre C. Press peer review of electronic search strategies: 2015 guideline statement. *J Clin Epidemiol* 2016;75:40–6.
- Barker TH, Migliavaca CB, Stein C, Colpani V, Falavigna M, Aromataris E, et al. Conducting proportional meta-analysis in different types of systematic reviews: A guide for synthesisers of evidence. *BMC Res Methodol* 2021;21(1):189.
- Hunter JP, Saratzis A, Sutton AJ, Boucher RH, Sayers RD, Bown MJ. In meta-analyses of proportion studies, funnel plots were found to be an inaccurate method of assessing publication bias. *J Clin Epidemiol* 2014;67(8):897–903.
- Borenstein M, Higgins JP, Hedges LV, Rothstein HR. Basics of meta-analysis: (I/2) is not an absolute measure of heterogeneity. *Res Synth Methods* 2017;8(1):5–18.
- Breman JG, Bernadou J, Nakano JH. Poxvirus in west african nonhuman primates: serological survey results. *Bull World Health Organ* 1977;55(5):605–12.
- Doty JB, Malekani JM, Kalembo LN, Stanley WT, Monroe BP, Nakazawa YU, et al. Assessing monkeypox virus prevalence in small mammals at the human-animal interface in the democratic republic of the congo. *Viruses* 2017;9(10).
- Douglas KO, Cayol C, Forbes KM, Samuels TA, Vapalahti O, Sironen T, et al. Serological evidence of multiple zoonotic viral infections among wild rodents in barbados. *Pathogens* 2021;10(6).
- Goldberg TL, Chapman CA, Cameron K, Saj T, Karesh WB, Wolfe ND, et al. Serologic evidence for novel poxvirus in endangered red colobus monkeys, western uganda. *Emerg Infect Dis* 2008;14(5):801–3.
- Guarner, et al. Monkeypox transmission and pathogenesis in prairie dogs. *Emerg Infect Dis* 2004;10(3):426–31.
- Hutin YJ, Williams RJ, Malfait P, Pebody R, Loparev VN, Ropp SL, et al. Outbreak of human monkeypox, democratic republic of congo, 1996 to 1997. *Emerg Infect Dis* 2001;7(3):434–8.
- Hutson CL, Lee KN, Abel J, Carroll DS, Montgomery JM, Olson VA, et al. Monkeypox zoonotic associations: Insights from laboratory evaluation of animals associated with the multi-state us outbreak. *Am J Trop Med Hyg* 2007;76(4):757–68.
- Marennikova SS, Seluhina EM, Mal'ceva NN, Ladnyj ID. Poxviruses isolated from clinically ill and asymptotically infected monkeys and a chimpanzee. *Bull World Health Organ* 1972;46(5):613–20.



- [50] Orba Y, Sasaki M, Yamaguchi H, Ishii A, Thomas Y, Ogawa H, et al. Orthopoxvirus infection among wildlife in zambia. *J Gen Virol* 2015;96(Pt 2):390–4.
- [51] Radonic A, Metzger S, Dabrowski PW, Couacy-Hymann E, Schuenadel L, Kurth A, et al. Fatal monkeypox in wild-living sooty mangabey, cote d'ivoire, 2012. *Emerg Infect Dis* 2014;20(6):1009–11.
- [52] Tee MS, Harrigan RJ, Thomassen HA, Smith TB. Ghosts of infections past: using archival samples to understand a century of monkeypox virus prevalence among host communities across space and time. *R Soc Open Sci* 2018;5(1):171089.
- [53] Bonilla-Aldana DK, García-Barco A, Jimenez-Diaz SD, Bonilla-Aldana JL, Cardona-Trujillo MC, Muñoz-Lara F, et al. Sars-cov-2 natural infection in animals: A systematic review of studies and case reports and series. *Vet Q* 2021;41(1):250–67.
- [54] Carrión-Nessi FS, Castro MP, Freitas-De Nobrega DC, Moncada-Ortega A, Omaña-Ávila ÓD, Mendoza-Millán DL, et al. Clinical-epidemiological characteristics and maternal-foetal outcomes in pregnant women hospitalised with covid-19 in venezuela: A retrospective study. *BMC Pregnancy Childbirth* 2022;22(1):905.
- [55] Amer F, Khalil HES, Elahmady M, ElBadawy NE, Zahran WA, Abdelnasser M, et al. Mpxv: Risks and approaches to prevention. *J Infect Public Health* 2023;16(6):901–10.
- [56] Amer FA, Hammad NM, Wegdan AA, ElBadawy NE, Pagliano P, Rodríguez-Morales AJ. Growing shreds of evidence for monkeypox to be a sexually transmitted infection. *Le Infez Med* 2022;30(3):323–7.
- [57] Abdelaal A, Reda A, Lashin BI, Katamesh BE, Brakat AM, Al-Manaseer BM, et al. Preventing the next pandemic: is live vaccine efficacious against monkeypox, or is there a need for killed virus and mrna vaccines? *Vaccines* 2022;10(9).
- [58] Shamim MA, Satapathy P, Padhi BK, Veeramachaneni SD, Akhtar N, Pradhan A, et al. Pharmacological treatment and vaccines in monkeypox virus: a narrative review and bibliometric analysis. *Front Pharm* 2023;14:1149909.
- [59] Katoto PD, Muttamba W, Bahizire E, Malembaka EB, Bosa HK, Kazadi DM, Lubambo G, Siangoli FB, Bakamutumaho B, Wayengera M, Mavungu Landu DJ, Mukadi-Bamuleka D, Mbala P, Loeb M, Kirenga B, Muyembe-Tamfum JJ. Shifting transmission patterns of human mpxv in South Kivu, DR Congo. *Lancet Infect Dis*. 2024 May 1:S1473-3099(24)00287-1. doi: 10.1016/S1473-3099(24)00287-1. Epub ahead of print. PMID: 38703785.
- [60] Mandja BA, Handschumacher P, Bompangue D, Gonzalez JP, Muyembe JJ, Sauleau EA, et al. Environmental drivers of monkeypox transmission in the democratic republic of the congo. *EcoHealth* 2022;19(3):354–64.
- [61] Tiwari A, Adhikari S, Kaya D, Islam MA, Malla B, Sherchan SP, et al. Monkeypox outbreak: Wastewater and environmental surveillance perspective. *Sci Total Environ* 2023;856(Pt 2):159166.
- [62] Hemati S, Mohammadi-Moghadam F. A systematic review on environmental perspectives of monkeypox virus. *Rev Environ Health* 2023.
- [63] Pozzetto B, Gagnaire J, Berthelot P, Bourlet T, Pillet S. [viruses present in the environment: virological considerations and examples of their impact on human health]. *Rev Francoph Des Lab: RFL* 2023;2023(550):33–43.
- [64] Wurtzer S, Levert M, Dhenain E, Boni M, Tournier JN, Londinsky N, et al. First detection of monkeypox virus genome in sewersheds in france: the potential of wastewater-based epidemiology for monitoring emerging disease. *Environ Sci Technol Lett* 2022;9(11):991–6.
- [65] Curaudeau M, Besombes C, Nakouné E, Fontanet A, Gessain A, Hassanin A. Identifying the most probable mammal reservoir hosts for monkeypox virus based on ecological niche comparisons. *Viruses* 2023;15(3).
- [66] Haider N, Guitian J, Simons D, Asogun D, Ansumana R, Honeyborne I, et al. Increased outbreaks of monkeypox highlight gaps in actual disease burden in sub-saharan africa and in animal reservoirs. *Int J Infect Dis: IJID: Publ Int Soc Infect Dis* 2022;122:107–11.
- [67] Rodríguez-Morales AJ, Ortiz-Martínez Y, Bonilla-Aldana DK. What has been researched about monkeypox? A bibliometric analysis of an old zoonotic virus causing global concern. *N Microbes N Infect* 2022;47:100993.
- [68] Reed KD, Melski JW, Graham MB, Regnery RL, Sotir MJ, Wegner MV, et al. The detection of monkeypox in humans in the western hemisphere. *N Engl J Med* 2004;350(4):342–50.
- [69] Di Giulio DB, Eckburg PB. Human monkeypox: an emerging zoonosis. *Lancet Infect Dis* 2004;4(1):15–25.
- [70] Shepherd W, Beard PM, Brookes SM, Frost A, Roberts H, Russell K, et al. The risk of reverse zoonotic transmission to pet animals during the current global monkeypox outbreak, united kingdom, june to mid-september 2022. *Eur Surveill* 2022;27(39).
- [71] Antinori S, Casalini G, Giacomelli A, Rodríguez-Morales AJ. Update on mpxv: a brief narrative review. *Infez Med* 2023;31(3):269–76.
- [72] Falendysz EA, Lopera JG, Lorenzonn F, Salzer JS, Hutson CL, Doty J, et al. Further assessment of monkeypox virus infection in gambian pouched rats (*Cricetomys gambianus*) using in vivo bioluminescent imaging. *PLoS Negl Trop Dis* 2015;9(10):e0004130.
- [73] Hutson CL, Carroll DS, Gallardo-Romero N, Drew C, Zaki SR, Nagy T, et al. Comparison of monkeypox virus clade kinetics and pathology within the prairie dog animal model using a serial sacrifice study design. *BioMed Res Int* 2015;2015:965710.
- [74] Abdelaal A, Serhan HA, Mahmoud MA, Rodríguez-Morales AJ, Sah R. Ophthalmic manifestations of monkeypox virus. *Eye (Lond)* 2023;37(3):383–5.
- [75] Abdelaal A, Reda A, Hassan AR, Mashaal A, Abu Serhan H, Katamesh BE, et al. Monkeypox-associated manifestations and complications involving the eye: a systematic review and meta-analysis of previous and current outbreaks. *Asia Pac J Ophthalmol (Philos)* 2023;12(3):326–37.
- [76] Sah R, Siddiq A, Reda A, Abdelaal A, Padhi BK, Mohanty A, et al. Oral manifestations of mpxv: a common presentation? *J Am Dent Assoc* 2023;154(4):279–82.
- [77] Sayad R, Siddiq A, Hashim A, Elsaedy AS. Can the current monkeypox affect the heart? a systematic review of case series and case report. *BMC Cardiovasc Disord* 2023;23(1):328.
- [78] Ortiz-Martínez Y, Zambrano-Sanchez G, Rodríguez-Morales AJ. Monkeypox and hiv/aids: when the outbreak faces the epidemic. *Int J STD AIDS* 2022;33(10):949–50.
- [79] Mitjà O, Alemany A, Marks M, Lezama Mora JI, Rodríguez-Aldama JC, Torres Silva MS, et al. Mpxv in people with advanced hiv infection: a global case series. *Lancet* 2023;401(10380):939–49.
- [80] Detecção de monkeypox em animal em minas gerais. <https://www.Saude.Mg.Gov.Br/component/gmg/story/17178-nota-informativa-sobre-deteccao-de-monkeypox-em-animal-em-minas-gerais-23-8-2022>.
- [81] Cimerman S, Chebabo A, Cunha CAD, Barbosa AN, Rodríguez-Morales AJ. Human monkeypox preparedness in latin america - are we ready for the next viral zoonotic disease outbreak after covid-19? *Braz J Infect Dis: Publ Braz Soc Infect Dis* 2022;26(3):102372.
- [82] Rodríguez-Morales AJ, Lopardo G, Verbanaz S, Orduna T, Lloveras S, Azeñas-Burgoa JM, et al. Latin america: situation and preparedness facing the multi-country human monkeypox outbreak. *Lancet Reg Health Am* 2022;13:100318.
- [83] Silva MST, Santos DGD, Coutinho C, Ribeiro MPD, Cardoso SW, Veloso VG, et al. The first case of acute hiv and monkeypox coinfection in latin america. *Braz J Infect Dis: Publ Braz Soc Infect Dis* 2023;27(2):102736.
- [84] Luz PM, Veloso VG, Grinsztejn B. The hiv epidemic in latin america: accomplishments and challenges on treatment and prevention. *Curr Opin HIV AIDS* 2019;14(5):366–73.
- [85] Silveira MF, Gomez Ponce de Leon R, Becerra F, Serruya SJ. Evolution towards the elimination of congenital syphilis in latin america and the caribbean: a multi-country analysis. *Rev Panam De Salud Publica = Pan Am J Public Health* 2019;43:e31.
- [86] Ortiz-Martínez Y, Saul Z, Hutchinson KA, Miljkovic G, Rodríguez-Morales AJ. Not just differential diagnoses... importance of sexually transmitted infections as coinfections with monkeypox amidst the outbreak. *Int J STD AIDS* 2022;33(13):1152–3.
- [87] Rodríguez-Morales AJ, León-Figueroa DA, Sah R, Villamil-Gomez WE. Arboviral diseases and monkeypox - an epidemiological overlapping differential diagnosis? *Rev Del Cuerpo Med Hosp Nac Almanzor Aguinaga Asenjo* 2022;15(3).
- [88] Chauhan RP, Fogel R, Limson J. Overview of diagnostic methods, disease prevalence and transmission of mpxv (formerly monkeypox) in humans and animal reservoirs. *Microorganisms* 2023;11(5).
- [89] Eteng WE, Mandra A, Doty J, Yinka-Ogunleye A, Aruna S, Reynolds MG, et al. Notes from the field: responding to an outbreak of monkeypox using the one health approach - nigeria, 2017-2018. *Mmwr Morb Mortal Wkly Rep* 2018;67(37):1040–1.
- [90] One health, monkeypox prevention, and treatment: The second online academic salon on monkeypox virus by the chinese association for laboratory animal sciences (august 26, 2022, beijing, china). *Anim models Exp Med* 2022;5(5):487–8.
- [91] Nuismer SL, Bull JJ. Self-disseminating vaccines to suppress zoonoses. *Nat Ecol Evol* 2020;4(9):1168–73.